

***Woessia lignicola* (Ramalinaceae, lichenized Ascomycota) a new combination and a new lichen species to Asia and Russia**

Yu. V. Storozhenko^{1,2}, E. A. Davydov¹, L. S. Yakovchenko³

¹Altai State University, Barnaul, Russia

²Tigirek State Nature Reserve, Barnaul, Russia

³Federal Scientific Center of East Asian Terrestrial Biodiversity of the Far Eastern Branch of the Russian Academy of Sciences, Vladivostok, Russia

Corresponding author: Yu. V. Storozhenko, yulia_storozhenko@mail.ru

Abstract. The epiphytic species *Woessia lignicola*, comb. nov., is newly reported for Asia and Russia from the Altai Territory (South Siberia). The species identity is supported by morphological examination and molecular-phylogenetic data (nrITS sequence). This finding extends the known distribution of the species eastward by more than 4000 km. In light of the recent resurrection of the genus *Woessia*, we propose the new combination *W. lignicola* (based on *Bacidina lignicola*).

Keywords: *Bacidina*, *Woessia*, biogeography, ITS, new record, taxonomy, Altai-Sayan Mountain system, Altai Territory, Salair Ridge.

Woessia lignicola (Ramalinaceae, lichenized Ascomycota) — новая комбинация и новый вид лишайника для Азии и России

Ю. В. Стороженко^{1,2}, Е. А. Давыдов¹, Л. С. Яковченко³

¹Алтайский государственный университет, Барнаул, Россия

²Государственный природный заповедник «Тигирекский», Барнаул, Россия

³Федеральный научный центр биоразнообразия наземной биоты Восточной Азии ДВО РАН, Владивосток, Россия

Автор для переписки: Ю. В. Стороженко, yulia_storozhenko@mail.ru

Резюме. Эпифитный вид *Woessia lignicola*, comb. nov., впервые обнаружен в Азии и России на территории Алтайского края (Южная Сибирь). Видовая принадлежность подтверждена морфолого-анатомическими и молекулярно-филогенетическими (последовательность nrITS) данными. Эта находка увеличивает известное распространение вида на восток более чем на 4000 км. В свете недавнего восстановления рода *Woessia* мы предлагаем новую комбинацию *W. lignicola* (на основе *Bacidina lignicola*).

Ключевые слова: *Bacidina*, *Woessia*, ITS, биоразнообразие, новые находки, таксономия, Алтае-Саянская горная страна, Алтайский край, Салаирский край.

Recent molecular phylogenetic studies have led to significant revisions within the lichen-forming family Ramalinaceae. Historically, the genus *Bacidina* Vězda was established to accommodate some species previously classified within the genera *Bacidia* De Not. and *Catillaria* A. Massal. (Vězda, 1991). The designated type species was *Bacidina phacodes* (Körb.) Vězda. Traditionally, *Bacidina* has been characterized by a thallus composed of goniocysts, a paraplechtenchymatous proper exciple, “*Lecanora*-type” asci, acicular or rarer more or less bacilliform ascospores, the absence of secondary metabolites, and filiform conidia (Vězda, 1983, 1991; Ekman, 2001; Kistenich *et al.*, 2018; Fatima *et al.*, 2021). Subsequent studies revealed considerable morphological diversity within the genus, including indistinct, granular, crustaceous, warted, areolate, microsquamulose to distinctly sorediate thallus (Ekman, 1996, 2023; Aptroot,

van Herk, 1999; Coppins, van den Boom, 2002; Brand *et al.*, 2009; Lumbsch *et al.*, 2011; Czarnota, Guzow-Krzemińska, 2012; Ekman *et al.*, 2012; Kondratyuk *et al.*, 2019; van den Boom, Alvarado, 2019; van den Boom, Magain, 2020; van den Boom, 2021). Additionally, molecular data showed *Bacidina* s. l. to be non-monophyletic (Kistenich *et al.*, 2018).

The genus *Woessia* D. Hawksw. et Poelt was recently resurrected based on robust multi-locus phylogenetic evidence (van den Boom, Alvarado, 2025). That study, utilizing sequences of mtSSU, LSU, and RPB1, recovered *Woessia* as a well-supported clade distinct from both *Bacidia* s. str. and *Bacidina* s. str. Consequently, several species previously classified in *Bacidina* were formally transferred to the resurrected genus *Woessia*, including its type species *W. fusarioides* Hawksw. et al. [current name *W. modesta* (Zwackh ex Vain.)

van den Boom et Alvarado] and other well-known taxa such as *W. arnoldiana* (Körb.) Sérus. et Die-derich, *W. chlorotricula* (Nyl.) Puntillo et al., and *W. egenula* (Nyl.) van den Boom et Alvarado (van den Boom, Alvarado, 2025).

Currently, the genera *Bacidina* and *Woessia* comprise approximately 60 worldwide-known species, 10 to 12 of which belong to the genus *Woessia* and have been newly described or resurrected in recent years (Fatima et al., 2021; Afshan et al., 2023; Ekman, 2023; van den Boom, Alvarado, 2025). Many of these recently recognized species have not been reported from Russia yet. One of the species recently described within the *Bacidina*/*Woessia* complex is *B. lignicola* S. Ekman known from dead wood in Scandinavia and Central Europe. Morphologically, it is very similar to *Woessia chlorotricula*, recently recorded from the Salair Ridge, Russia (Kotkova et al., 2022), and to *Bacidina phacodes*. During a revision of Russian material of these morphologically similar species, we identified specimen matching the description of *B. lignicola*. Given the reinstatement of the genus *Woessia*, the taxonomic placement of this species requires revision. In this paper we (1) confirm the first record of *Bacidina lignicola* in Asia and Russia based on morphological and molecular (nrITS) data from the Altai Territory, (2) formally transfer *B. lignicola* to the genus *Woessia* to align its nomenclature with the current phylogenetic framework.

Material and Methods

Study area and material collection. The material was collected by the authors in Salair National Park on the Salair Ridge in the Altai Territory. The Salair Ridge is a low-mountain area within the northwest of the Altai-Sayan Mountain system with an area of 22500 km² with average elevations of 400–600 m a. s. l. and the highest mountain Mt. Kivda, 621 m a. s. l. (Lashchinskii et al., 2007; Bykov et al., 2021).

Morphological studies. The specimens were examined using a stereomicroscope (Zeiss Stemi 2000-C) and a compound microscope (Zeiss Axio Lab.A1). Anatomical examination was undertaken using hand-cut sections mounted in water. Measurements of morphological characters follow Vondrák et al. (2013), morphological terminology follows Ekman (2023).

Molecular studies. DNA extraction, amplification and sequencing followed the methods described by Davydov and Yakovchenko (2017). The primer sets were ITS1F (Gardes, Bruns, 1993) and

ITS4 (White et al., 1990) The program Geneious 6.0 (Biomatters Ltd., New Zealand) was used for assembling sequences. Consensus sequences were exclusively compiled from double-stranded parts of the sequences.

The newly generated nrITS sequence of *Woessia lignicola* was supplemented with nrITS and mtSSU sequences related to *Woessia* and morphologically close *Bacidina* species referable from the phylogenetic studies of *Bacidina* s. l. (Vondrák et al., 2023, 2024; van den Boom, Alvarado, 2025) obtained from GenBank (Table 1). *Bellicidia incompta* (Borrer) Kistenich et al. was designated as the outgroup, because in the recent phylogenetic studies *Bellicidia* is closely related to *Bacidia* and *Bacidina* (e. g., Kistenich et al., 2018).

Sequence alignment was performed using the MAFFT algorithm (Kato et al., 2005) on the GUIDANCE web server (Sela et al., 2015) with default settings. The aligned dataset was partitioned by locus ITS1, 5.8S, ITS2, and mtSSU, with each partition treated independently in the subsequent analysis.

Maximum likelihood (ML) analysis was conducted using RAXML 8.0.26 (Stamatakis, 2014) via raxmlGUI v.1.3.1 (Silvestro, Michalak, 2012). The GTRGAMMA model of evolution was applied to each partition. Branch support was assessed with 1000 rapid bootstrap replicates, and a majority-rule consensus tree was generated.

Results and Discussion

Phylogenetic placement and confirmation of identity. Phylogenetic reconstruction based on the combined dataset of nrITS and mtSSU sequences (Fig. 1) resolves four main clades with high statistical support: 1) a clade corresponding to the genus *Woessia*, including the type species *W. modesta* (82% BS); 2) a clade uniting *Bacidina violacea* and *B. populnea* (99% BS); 3) a clade corresponding to the genus *Bacidina*; and 4) a clade corresponding to the genus *Bacidia*. In addition to the confirmed members of *Woessia*, the *Woessia* clade includes *Bacidina assulata*, *B. lignicola*, *B. paradoxa*, and two putatively new *Bacidina* s. l. species (*Bacidina* sp. 2 and 3). *Bacidina adastrata* and *B. pycnidiata* likely also belong to *Woessia* (Fig. 1). While a comprehensive taxonomic revision of all these taxa is beyond the scope of the present study and requires further investigation, the close morphological affinity of *Bacidina lignicola* to *Woessia chlorotricula* prompted us to address its nomenclatural status. Consequently,

Table 1

Collection and GenBank accession numbers of the specimens used in the phylogenetic analysis in this study

Species and № in phylogenetic tree	Specimen numbers	GenBank accession numbers	
		ITS	mtSSU
<i>Bacidia abisistens</i>	BG, Ekman 3223	AF282085	MG925845
<i>B. arceutina</i>	PRA, Vondrák 24590	OQ717709	OQ646107
<i>B. caucasica</i>	GLM, Otte 0048447	MW523553	MW506388
<i>B. laurocerasi</i>	PRA, Vondrák 22972	OQ717714	OQ646109
<i>B. polychroa</i>	PRA, Vondrák 26081	OQ717313	OQ682869
<i>B. rosella</i>	PRA, Vondrák 26028	OQ717314	OQ682870
<i>B. rubella</i>	PRA, Vondrák 26027	OQ717316	OQ682871
<i>Bacidina acerina</i> 1	PRA, Vondrák 24355	OK332880	OK465494
<i>B. acerina</i> 2	PRA, Vondrák 20656	OK332878	OK465492
<i>B. adastr</i> 1	PRA, Vondrák 24569	OK332938	-
<i>B. adastr</i> 2	Herb. Malíček 13928	OQ717318	-
<i>B. assulata</i> 1	PRA, Palice 30714	OQ717321	-
<i>B. assulata</i> 2	PRA, Vondrák 24106	OQ717721	OQ646116
<i>B. assulata</i> 3	PRA, Vondrák 24102	OQ717722	OQ646115
<i>B. caerulea</i>	BG, Ekman 3579	OP256853	-
<i>B. friesiana</i>	BG, Ekman 3401	AF282101	-
<i>B. paradoxa</i> 1	PRA, Palice 26142	OQ717326	OQ682876
<i>B. paradoxa</i> 2	PRA, Palice 33646	OQ717327	OQ682877
<i>B. phacodes</i> 1	PRA, Vondrák 25994	OQ717322	-
<i>B. phacodes</i> 2	PRA, Vondrák 24463	OQ717723	-
<i>B. piceae</i>	PRA, Palice 32287	OQ717724	OQ646117
<i>B. populnea</i>	UPS, L-1009299	OP256855	-
<i>B. pycnidia</i>	Herb. Malíček 2553	KX239034	-
<i>B. violacea</i>	PRA, Palice 31173	OQ717328	-
<i>Bacidina</i> sp. 2 (1)	Herb. Malíček 15981	OQ918723	OQ920110
<i>Bacidina</i> sp. 2 (2)	PRA, Vondrák 24921	OK332885	OK465501
<i>Bacidina</i> sp. 3	PRA, Vondrák 24573	OK332886	OK465503
<i>Bellicidia incompta</i>	PRA, Vondrák 23534	OQ717713	OQ646108
<i>Woessia arnoldiana</i> 1	PRA, Vondrák 24753	OQ717717	OQ646112
<i>W. arnoldiana</i> 2	BG, Ekman 3157	-	MG925854
<i>W. brandii</i>	GPN, Czarnota 7599	KX239029	-
<i>W. chlorotricula</i> 1	PRA, Palice 32914	OQ717320	-
<i>W. chlorotricula</i> 2	B, Sipman 55079	-	JQ796833
<i>W. chlorotricula</i> 3	PRA, Vondrák 26080	OQ717319	OQ682874
<i>W. delicata</i>	Sérusiaux s. n. (LG, DNA 369)	JQ796854	JQ796834
<i>W. egenula</i>	LG, van den Boom 39665	-	JQ796836
<i>W. inundata</i>	BG, Ekman 3187	-	MG925855
<i>W. lignicola</i> 1	PRA, Vondrák 24312	OQ717719	OQ646113
<i>W. lignicola</i> 2	PRA, Vondrák 25879	OQ717720	OQ646114
<i>W. lignicola</i> 3	ALTB, Davydov 23847, Storozhenko	PZ103500	-
<i>W. lignicola</i> 4	UPS-L, Ekman 941645	OP256857	-
<i>W. modesta</i> 1 (type)	DUKE-AFTOL-ID 1845	-	DQ986810
<i>W. modesta</i> 2	GPN, Czarnota 7246	JN972447	-
<i>W. modesta</i> 3	GPN, Czarnota 5275	JN972445	-
<i>W. neosquamulosa</i> 1	UPS, van den Boom 24182	OP256854	-
<i>W. neosquamulosa</i> 2	LG, van den Boom 39692	-	JQ796838
<i>W. terricola</i>	BR, van den Boom 55461	-	MW622008

Note: the newly generated sequence is in bold.

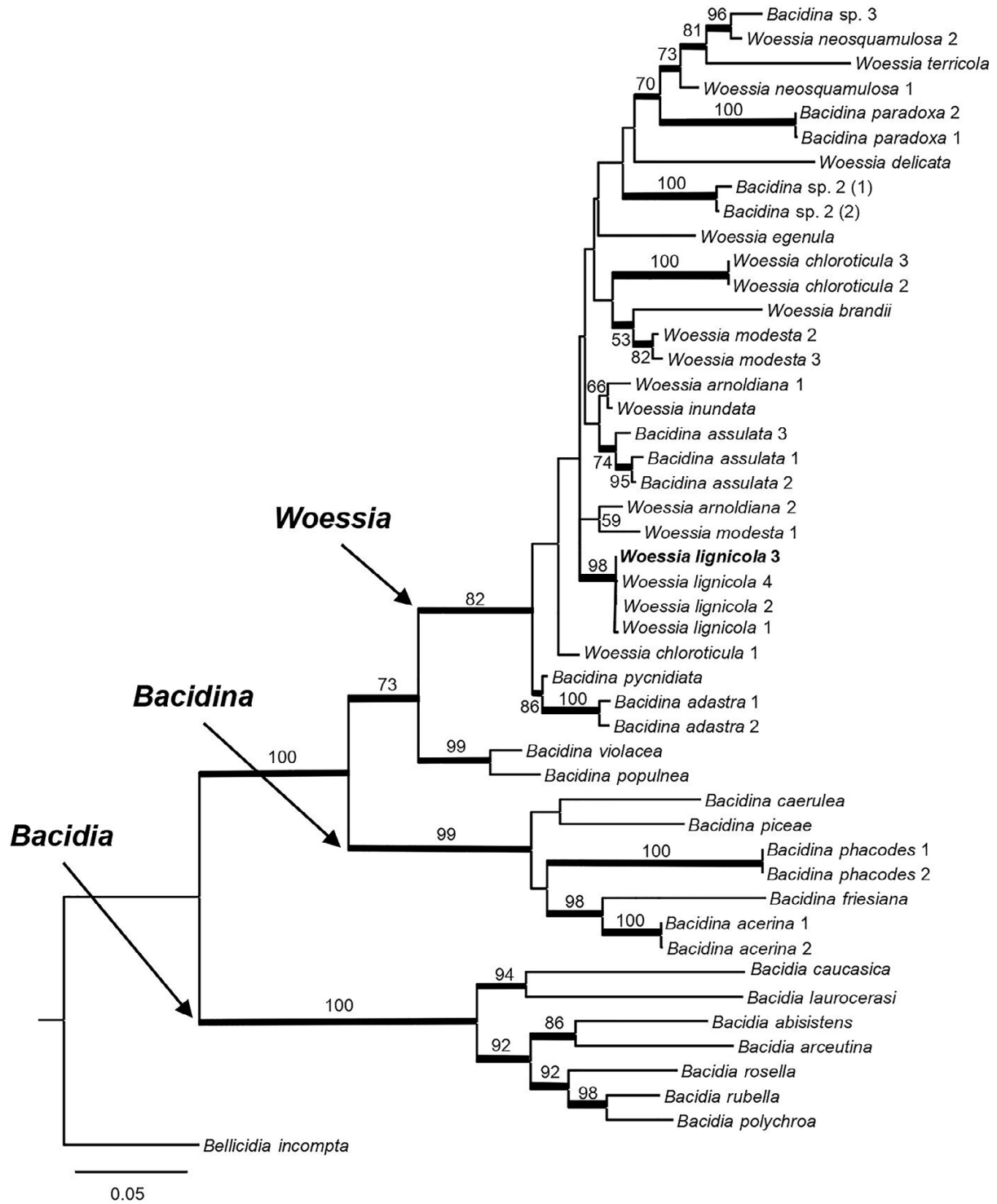


Fig. 1. Phylogenetic relationships among *Woessia* and *Bacidina* s. l. species based on a ML + rapid bootstrap combined analysis of ITS1-5.8S-ITS2 and mtSSU. Numbers on branches represent maximum likelihood bootstrap values (BS; $\geq 70\%$ is bold). The newly generated sequence from Russia is highlighted in bold. Corresponding GenBank accession numbers are provided in Table 1.

we propose the new combination *Woessia lignicola* below.

Within the *Woessia* clade, the newly generated sequence from Russia (PZ103500) groups with high statistical support (BS = 98) with reference sequences from the type material of *W. lignicola*

from Sweden (OP256857) and the Czech Republic (OQ717719, OQ717720), confirming the identity of our specimen as *W. lignicola*. According to the distance matrix, the Russian ITS sequence differs from the Swedish sequence by a single nucleotide and is identical to the Czech sequences.

Morphological distinction from similar species. *Woessia lignicola* has previously been confused with *Bacidina phacodes* and *Woessia chlorotricula* due to superficial similarities in apothecium size, coloration, and overlapping substrates. However, detailed anatomical examination reveals consistent differences in thallus structure, apothecia, proper exciple, and conidia, allowing for reliable separation of these species (Ekman, 2023). While *W. lignicola* and *Bacidina phacodes* share a similar anatomical structure, the main differences lie in apothecial shape and hymenium height. *Woessia lignicola* and *W. chlorotricula* can be distinguished by differences in the external and internal structure of their apothecia (Ekman, 2023). Key diagnostic characters are summarized in Table 2.

The examined specimens of *Woessia lignicola* and *W. chlorotricula* clearly differ in apothecial color, the structure of the apothecial margin, and the thickness of the proper exciple. The Russian specimen of *W. lignicola* (Fig. 2) is characterized by pale pinkish to pale orange apothecia, a hymenium 37–43 μm tall, constituting 25–40% of the total apothecium height, that does not correspond to the known literature (Ekman, 2023),

in which the height of the hymenium is 10–20%, a distinctly radiating proper exciple, and colorless, acicular, 0–5–7-septate, straight ascospores 25.0–30.0–42.0 $\times$ 1.2–1.4–1.7 μm ($n = 20$). Pycnidia were not detected.

Specimens examined. *Woessia lignicola*: **Russia, Altai Territory**: Togul'skii District, Salair Ridge, headwaters of the Malaya Mostovaya River at 10 km NEE from the Novokamenka settlement, 53°23'28"N, 86°27'39"E, 400 m a. s. l., *Abies sibirica* Ledeb. dominated subnemoral forest, on fruit body of polyporoid fungus on bark of *Populus tremula* L., 15 IX 2022, Davydov 23847, Storozhenko, ALTB.

Woessia chlorotricula: **Russia, Altai Territory**: Zarin'skii District, Salair Ridge, at 13 km to NE from the Novoiushino, Togul River basin, 53°42'07"N, 85°59'44"E, 335 m a. s. l., linden (*Tilia sibirica* Bayer) forest, on bark of *Padus avium* L., 22 VIII 2020, Davydov 21314, Storozhenko, det. Storozhenko, Yakovchenko, Davydov, ALTB; *ibid.*, 53°41'41.1"N, 86°00'05.8"E, 315 m a. s. l., fir-poplar (*Abies sibirica*-*Populus tremula*) subnemoral forest, on fruit body of polyporoid fungus, 23 VIII 2020, Davydov 21306, Storozhenko, ALTB.

Bacidina phacodes: **Russia, Altai Territory**: Togul'skii District, Salair Ridge, at 13 km to NE from the Novoiushino, Togul River basin, 53°42'03.9"N, 85°59'38.3"E, 330 m a. s. l., linden (*Tilia sibirica*) forest, on bark of *Populus tremula*, 24 VIII 2020, Davydov 21730, Storozhenko, ALTB.

Table 2

Key diagnostic characters of similar *Woessia* and *Bacidina* species

Character	<i>Woessia lignicola</i>	<i>Bacidina phacodes</i>	<i>Woessia chlorotricula</i>
1. Substrate and thallus	Primarily fresh stumps of conifers (<i>Picea abies</i> , <i>Pinus sylvestris</i>) and hard wood; thallus thin or thick, of scattered convex areoles or continuous and warted	Soft bark of deciduous trees; thallus thin to irregularly warted or scurfy, confined to bark crevices	Diverse substrates: wood, bark, rock, metal, moribund bryophytes; thallus thin, filmy or of discrete granules
2. Apothecia: size, shape, color	0.15–0.44 mm diam, convex, pale pinkish (becoming brownish-orange in herbarium)	Less than 0.4 mm diam., effuse, pale pink	0.15–0.25 mm diam., flat, pale greyish yellow or pale brown-grey
3. Apothecial margin	Usually slightly paler than disk, smooth, soon excluded	Paler than disk, smooth, soon excluded	Slightly paler than disc, almost hairy, persistent
4. Hymenium and hypothecium height	Hymenium makes up 10–20% (Ekman, 2023) and 25–40% (for author's specimen) of apothecium height, mostly colorless; thick hyphal cushion below hypothecium. Hypothecium colorless	Hymenium makes up 30–40% of apothecium height, mostly colorless; no hyphal cushion. Hypothecium colorless	Hymenium relatively tall (35–50% of apothecium height), mostly colorless, without special thickening. Hypothecium pale
5. Proper exciple structure	Excipular hyphae distinctly radiating, cell lumina up to 12 $\times$ 4 μm	Excipular hyphae weakly radiating; cell lumina up to 7.0 $\times$ 3.3 μm	Exciple paraplectenchymatous, hyphae not radiating, with vesicle-like lumina, cell lumina up to 7 $\times$ 4 μm
6. Conidia	Acicular, straight, non-septate or weakly 1–3-septate, 22.0–43.0 $\times$ 1.2 μm	Acicular, straight, resembling ascospores, 3–7-septate, (24)28–50(60) $\times$ 1.0–1.5(2.0) μm	Mostly filiform, occasionally pyriform, curved, non-septate, 19.0–48.0 $\times$ 0.8–1.0 μm
7. Pycnidia	Abundant, 50–130 μm diam., immersed to semi-immersed	Sparse, 120–200 μm diam., immersed	Sparse, 40–90 μm diam., immersed or semi-immersed

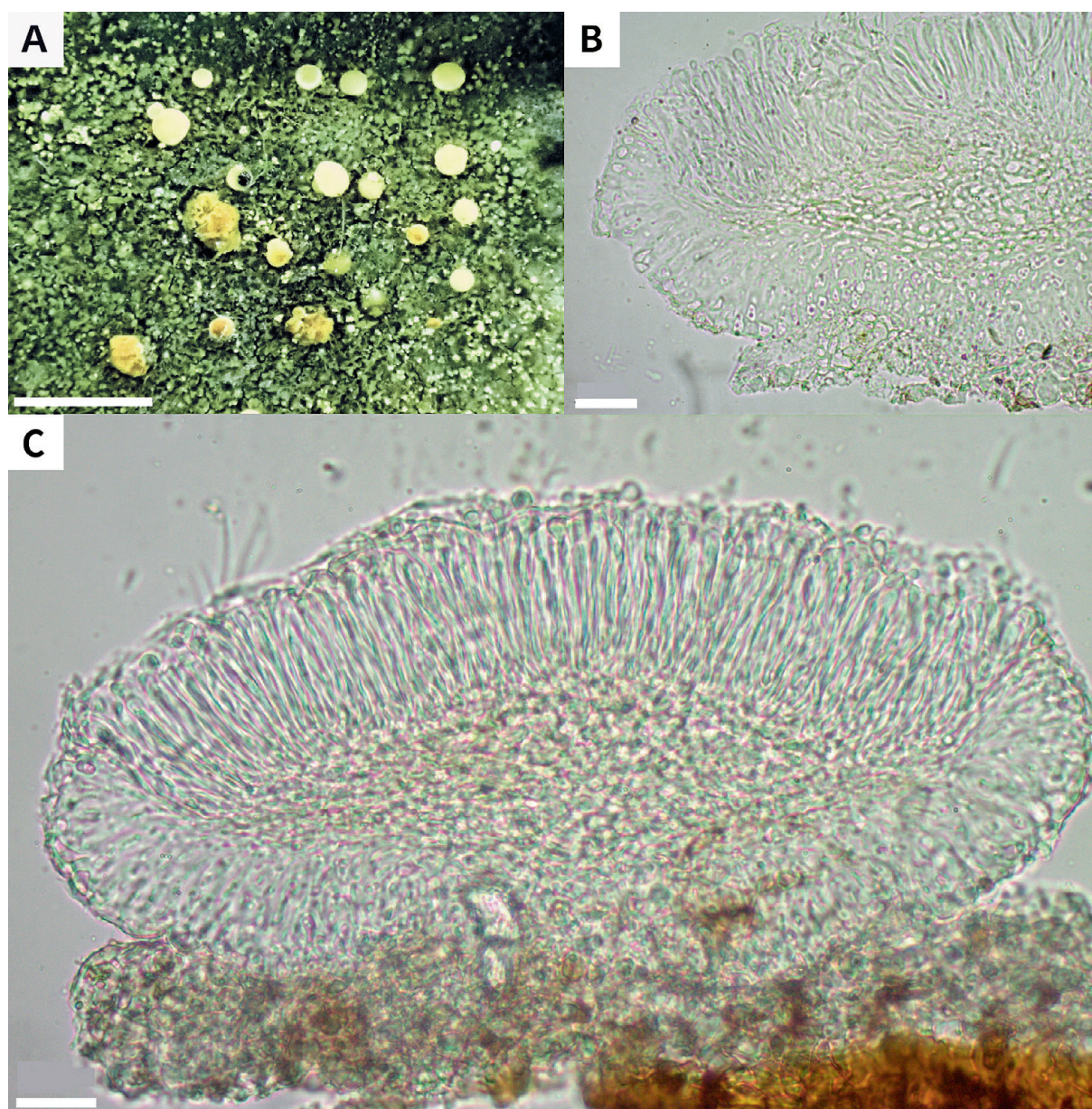


Fig. 2. Russian specimen of *Woessia lignicola* (ALTB, Davydov 23847, Storozhenko): A — habitus; B — anatomical structure of the proper exciple; C — apothecial structure. Scale bars: A — 1 mm, B, C — 20 µm.

Distribution and ecology. *Woessia lignicola* was previously known only from scattered localities in Northern and Central Europe — Norway, Finland, Poland, Germany and the Czech Republic, where it occurs primarily on recently dead wood of conifers and hardwoods in humid, shaded forests (Ekman, 2023). Our discovery in the Salair Ridge (the Altai Territory) represents a significant eastward range extension of more than 4000 km, marking the first record for Asia and Russia. Here the species was found in a low-mountain area within the Altai-Sayan system in old-growth mixed forests. This suggests that *W. lignicola* may have a broader boreal distribution across Eurasia, possibly linked to relict forest refugia. The disjunct distribution pattern resembles that of several other

epiphytic lichens with European-Siberian ranges (Davydov, Printzen, 2012; Davydov, Konoreva, 2017), highlighting the importance of South Siberia and particularly Altai as a biogeographic corridor and refugium for temperate and boreal lichen species (Davydov, Printzen, 2012). Further studies in Siberia and the Russian Far East are needed to clarify full distribution of *W. lignicola*.

Taxonomy. In accordance with the revised generic delineation within the Ramalinaceae proposed by van den Boom, Alvarado (2025), which recognizes *Woessia* as a distinct genus, we formally transfer *Bacidina lignicola* to the genus *Woessia*. The new combination is proposed below.

Woessia lignicola (S. Ekman) Storozhenko, Davydov et Yakovchenko, comb. nov.

MycoBank No.: MB862208

Basionym: *Bacidina lignicola* S. Ekman, Nordic Journal of Botany 2023 (e03846): 12 (2023).

Type: Sweden, Gästrikland, Torsåker par., Hofors herrgård, the park S of the manor, 50°33'05.7"N, 16°17'50.0"E, 120 m a. s. l., on *Ulmus* stump, 23 IX 2018, S. Ekman 5749, UPS L-971089 holotype, not seen.

Acknowledgments

The study of Yulia Storozhenko was carried out as a part of the implementation of the Altai State University Development Program for 2021–2030 as a part of the implementation of the strategic academic leadership program “Priority 2030”, project Assessment of genetic diversity of epiphytic lichens in the Salair National Park. The research of Lidia Yakovchenko was carried out within the state assignment of Ministry of Science and Higher Education of the Russian Federation (theme No. 124012400285-7).

Contribution of the authors. Yu. V. Storozhenko – collection and study of samples, data analysis, writing; E. A. Davydov – organization of expedition, collection of samples, methodology, data analysis, writing; L. S. Yakovchenko – methodology, sample examination, data analysis. All the authors have read and agreed with the final submitted version of the manuscript.

Conflicts of Interest. The authors declare no conflict of interest.

References / Литература

- Afshan N., Fayyaz I., Iftikhar F., Niazi A. R., Habib K., Khalid A. N. 2023. A new species of the genus *Bacidina* (Ramalinaceae, Ascomycota) from Pakistan. *Herzogia* 36(2): 470–478. <https://doi.org/10.13158/heia.36.2.2023.470>
- Aptroot A., van Herk C. M. 1999. *Bacidia neosquamulosa*, a new and rapidly spreading corticolous lichen species from Western Europe. *The Lichenologist* 31(2): 121–127. <https://doi.org/10.1006/lich.1998.0184>
- Brand M., Coppins B., van den Boom P. P. G., Sérusiaux E. 2009. Further data on the lichen genus *Bacidia* s. l. in the Canary Islands and Western Europe, with descriptions of two new species. *Bibliotheca Lichenologica* 99: 81–92.
- Bykov N. I., Vazhov S. V., Garms O. Ya., Gribkov A. V., Davydov E. A., Elesova N. V., Zyablintseva M. V., Kosachev P. A., Kuzmenkin D. V., Maslova O. M. et al. 2021. Natural conditions of the Salair National Park. *Proceedings of the Tigirek State Natural Reserve* 13: 7–45. [Быков Н. И., Важов С. В., Гармс О. Я., Грибков А. В., Давыдов Е. А., Елесова Н. В., Зяблинцева М. В., Косачев П. А., Кузменкин Д. В., Маслова О. М. и др. 2021. Природные условия национального парка «Салаир». *Труды Тигирекского заповедника* 13: 7–45].
- Coppins B. J., van den Boom P. P. G. 2002. *Bacidia brandii*, a new lichen species from the Netherlands, Belgium, France and Lithuania. *The Lichenologist* 34(4): 327–332. <https://doi.org/10.1006/lich.2002.0403>
- Czarnota P., Guzow-Krzemińska B. 2012. ITS rDNA data confirm a delimitation of *Bacidina arnoldiana* and *B. sulphurella* and support a description of a new species within the genus *Bacidina*. *The Lichenologist* 44(6): 743–755. <https://doi.org/10.1017/S0024282912000515>
- Davydov E. A., Konoreva L. A. 2017. New data on lichens from Salair province in Altaisky krai (Siberia, Russia). *Turczaninowia* 20(4): 185–197. <https://doi.org/10.14258/turczaninowia.20.4.17>
- Davydov E. A., Printzen C. 2012. Rare and noteworthy boreal lichens in Altai Mountains (South Siberia, Russia). *The Bryologist* 115(1): 61–73. <https://doi.org/10.2307/41486742>
- Davydov E. A., Yakovchenko L. S. 2017. *Rhizocarpon smaragdulum*, a new monosporic yellow-thalline species and some additional species of the genus *Rhizocarpon* from the Altai Mountains (Siberia). *The Lichenologist* 49(5): 457–466. <https://doi.org/10.1017/S0024282917000469>
- Ekman S. 1996. The corticolous and lignicolous species of *Bacidia* and *Bacidina* in North America. *Opera Botanica* 127: 1–148.
- Ekman S. 2001. Molecular phylogeny of the Bacidia-ceae (Lecanorales, lichenized Ascomycota). *Mycological Research* 105: 783–797. <https://doi.org/10.1017/S0953756201004269>
- Ekman S. 2023. Four new and two resurrected species of *Bacidina* from Sweden, with notes and a preliminary key to the known Scandinavian species. *Nordic Journal of Botany* 2023(5): e03846. <https://doi.org/10.1111/njb.03846>
- Ekman S., Jonsson F., Hermansson J. 2012. *Bacidina etayana* and *B. saxenii* new to Sweden. *Graphis Scripta* 24(1): 14–18.
- Fatima M., Habib K., Czarnota P., Khalid A. N. 2021. Two new *Bacidina* species (Lecanorales, Ascomycota) from Pakistan. *Folia Cryptogamica Estonica* 58: 25–34. <https://doi.org/10.12697/fce.2020.58.03>
- Gardes M., Bruns T. D. 1993. ITS primers with enhanced specificity for basidiomycetes – application to the identification of mycorrhizae and rusts. *Molecular Ecology* 2: 113–118. <https://doi.org/10.1111/j.1365-294X.1993.tb00005.x>
- Katoh K., Kuma K., Toh H., Miyata T. 2005. MAFFT version 5: improvement in accuracy of multiple sequence alignment. *Nucleic Acids Research* 33: 511–518. <https://doi.org/10.1093/nar/gki198>
- Kistenich S., Timdal E., Bendiksby M., Ekman S. 2018. Molecular systematics and character evolution in the lichen family Ramalinaceae (Ascomycota: Lecanorales). *Taxon* 67: 871–904. <https://doi.org/10.12705/675>
- Kondratyuk S. Y., Lőkös L., Farkas E., Jang S.-H., Liu D., Halda J., Persson P.-E., Hansson M., Kärnefelt I., Thell A. et al. 2019. New and noteworthy lichen-forming and lichenicolous fungi 9. *Acta Botanica Hungarica* 61(3–4): 325–367. <https://doi.org/10.1556/034.61.2019.3-4.6>
- Kotkova V. M., Afonina O. M., Androsova V. I., Arslanov S. N., Belyakov E. A., Chernova A. M., Czernyadjeva I. V., Davydov E. A., Doroshina G. Ya., Erokhina O. V. et al. 2022. New cryptogamic records. 10. *Novosti sistematiki nizshikh rastenii* 56(2): 477–517. <https://doi.org/10.31111/nsnr/2022.56.2.477>
- Lashchinskii N. N., Sedel'nikova N. V., Safonova T. A., Pisarenko O. Yu., Perova N. V., Gorbunova I. A.,

- Lashchinskaya N. V. 2007. *Flora Salairskogo kryazha* [Flora of the Salair Ridge]. Novosibirsk: 252 p. [Лашчинский Н. Н., Седельникова Н. В., Сафонова Т. А., Писаренко О. Ю., Перова Н. В., Горбунова И. А., Лашчинская Н. В. 2007. *Флора Салаирского кряжа*. Новосибирск: 252 с.].
- Lumbsch H. T., Ahti T., Altermann S., De Paz G. A., Aptroot A., Arup U., Lücking R. 2011. One hundred new species of lichenized fungi: a signature of undiscovered global diversity. *Phytotaxa* 18(1): 1–127. <https://doi.org/10.11646/phytotaxa.18.1.1>
- Sela I., Ashkenazy H., Katoh K., Pupko T. 2015. GUID-ANCE2: accurate detection of unreliable alignment regions accounting for the uncertainty of multiple parameters. *Nucleic Acids Research* 43: W7–W1. <https://doi.org/10.1093/nar/gkv318>
- Silvestro D., Michalak I. 2012. raxmlGUI: a graphical front-end for RAxML. *Organism Diversity and Evolution* 12: 335–337. <https://doi.org/10.1007/s13127-011-0056-0>
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30: 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- van den Boom P. P. G. 2021. Follicolous lichens and their lichenicolous fungi in Macaronesia and atlantic Europe. *Bibliotheca Lichenologica* 111: 1–197. <https://doi.org/10.1017/S0024282922000287>
- van den Boom P. P. G., Alvarado P. 2019. Lichens and lichenicolous fungi of Faial (Azores, Portugal) with descriptions of three new species. *Herzogia* 32(2): 421–437. <https://doi.org/10.13158/heia.32.2.2019.421>
- van den Boom P. P. G., Alvarado P. 2025. New species of *Bacidia* s. l. from the Azores and the resurrection of genus *Woessia*. *Diversity* 17(3): 187. <https://doi.org/10.3390/d17030187>
- van den Boom P. P. G., Magain N. 2020. Three new lichen species from Macaronesia belonging in Ramalinaceae, with the description of a new genus. *Plant and Fungal Systematics* 65(1): 167–175. <https://doi.org/10.35535/pfsyst-2020-0011>
- Vězda A. 1983. Foliicole Flechten aus der Kolchis (West-Transkaukasien, UdSSR). *Folia Geobotanica Phytotaxonomica* 18: 45–70. <https://doi.org/10.1007/BF02855637>
- Vězda A. 1991 (1990). *Bacidina* genus novum familiae Lecideaceae (Ascomycetes lichenisati). *Folia Geobotanica Phytotaxonomica* 25: 431–432. <https://doi.org/10.1007/BF02914011>
- Vondrák J., Frolov I., Arup U., Khodosovtsev A. 2013. Methods for phenotypic evaluation of crustose lichens with emphasis on Teloschistaceae. *Chornomorskiy Botanichniy Zhurnal* 9(3): 382–405. <https://doi.org/10.14255/2308-9628/13.93/>
- Vondrák J., Svoboda S., Košnar J., Malíček J., Šoun J., Frolov I., Svensson M., Novotný P., Palice Z. 2023. Martin7: a reference database of DNA barcodes for European epiphytic lichens and its taxonomic implications. *Preslia* 95(3): 311–345. <https://doi.org/10.23855/preslia.2023.311>
- Vondrák J., Svoboda S., Malíček J., Šoun J., Košnar J., Svensson M., Timdal E., Macháč J., Palice Z. 2024. Combining environmental DNA data and taxonomic surveys provides an unprecedented understanding of lichen diversity and accelerates the discovery of new species. *Preslia* 96(4): 351–417. <https://doi.org/10.23855/preslia.2024.351>
- White T. J., Bruns T., Lee S., Taylor J. W. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR protocols: a guide to methods and applications*. New York: 315–322. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>