Worldwide genetic relationships of pigs as inferred from X chromosome SNPs


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Summary

The phylogeography of the porcine X chromosome has not been studied despite the unique characteristics of this chromosome. Here, we genotyped 59 single nucleotide polymorphisms (SNPs) in 312 pigs from around the world, representing 39 domestic breeds and wild boars in 30 countries. Overall, widespread commercial breeds showed the highest heterozygosity values, followed by African and American populations. Structuring, as inferred from $F_{ST}$ and analysis of molecular variance, was consistently larger in the non-pseudoautosomal (NPAR) than in the pseudoautosomal regions (PAR). Our results show that genetic relationships between populations can vary widely between the NPAR and the PAR, underscoring the fact that their genetic trajectories can be quite different. NPAR showed an increased commercial-like genetic component relative to the PAR, probably because human selection processes to obtain individuals with high productive parameters were mediated by introgressing boars rather than sows.

Keywords animal genetic resources, phylogeography, pig, sex chromosome, single-nucleotide polymorphism, wild boar.

Introduction

Archeological and genetic evidence shows that the pig was repeatedly domesticated from wild boars starting in the Neolithic period, initially in eastern Asia and later in other regions of Asia and of Europe (Larson et al. 2005; Fang & Andersson 2006). Our knowledge of the origin of the different pig populations subsequent to domestication is still incomplete. In Asia, differences between northern and southern wild boar populations have been found, and multiple ancestors for domesticated Asian pig populations have been reported (Fang et al. 2005; Luetkemeier et al. 2010). In Europe, the wild boar was domesticated in the Neolithic period in different places and eventually formed European local breeds (Giuffra et al. 2000; Ramirez et al. 2009). An introgression of Asian pigs into European breeds during the 18th and 19th centuries is well documented (Porter 1993; Giuffra et al. 2000). This event resulted in today’s commercial European breeds, most of which are now widespread internationally. Other populations have been much less widely studied. In the Americas, the species was introduced during Columbus’s second trip to the Hispaniola Island (today, Dominican Republic and Haiti) in 1493. The current main genetic component of these animals is supposed to be Iberian, with a possible subsequent introgression of other breeds. As for African populations, Ramirez et al. (2009) reported an Asian component in pigs in East African countries that was not observed in the western part of the continent.

Several approaches have been used to study population genetic relationships in pigs, including mtDNA sequences (Giuffra et al. 2000; Larson et al. 2005; Fang & Andersson 2006), the Y chromosome (Ramirez et al. 2009), and
autosomal genetic markers (Fan et al. 2002; Fang et al. 2005; SanCristobal et al. 2006). In contrast, X chromosome markers have been neglected so far. Yet, some characteristics make the X chromosome an interesting resource for the study of variability between populations (Schaffner 2004). Despite the structural differences between the X and Y chromosomes, they have a region that allows for segregation in meiosis, called the pseudoautosomal region (PAR). This region, located in the telomeric regions of the X chromosome arms, can recombine with the Y chromosome (Schaffner 2004). The rest of the chromosome constitutes the non-pseudoautosomal region (NPAR). Males carry only one copy of this chromosome, making its effective population size three-quarters that of the autosomes and increasing genetic drift for the NPAR-linked loci. Also, differences between effective population sizes in each gender can influence differences between populations depending on the migration sex ratio (Pool & Nielsen 2009; Casto et al. 2010).

The PAR X chromosome structure has been described in bovine (Das et al. 2009), horse (Raudsepp & Chowdhary 2008), dog and sheep (Toder et al. 1997). In pigs, the first PAR genes (PRKX, KAL1, and STS) were mapped to SSCXp/Yp by Quilter et al. (2002). Also, the porcine pseudoautosomal boundary (PAB) was recently mapped by qPCR between SHROOM2 and CLCN4 (Raudsepp et al. 2012). According to the Sscrofa9 assembly (Archibald et al. 2010), the Sus scrofa X chromosome (SSCX) is 125.8 Mb long and contains 701 coding genes, 32 pseudogenes, and 178 non-coding RNAs (www.ensembl.org).

Here, we report on the analysis of worldwide pig population relationships using SSCX SNP polymorphisms in both the NPAR and the PAR, and we refine the localization of both regions in the porcine genome.

**Materials and methods**

**Samples**

A total of 312 DNA samples from 224 males, 87 females, and one unknown-sex individual from 39 pig breeds collected in 30 countries around the world were genotyped in this study (Table S1). These breeds include local breeds and wild boars from Asia and Europe, local creole and feral pigs from the Americas, and a smaller sample from African countries. Information about breed was not available for the latter region. Within commercial breeds, we included the most widely used breeds in the modern pig industry, mostly of European origin, as well as some British local breeds that have been introgressed with Asian germplasm (Table S1).

**SNP selection, genotyping, and quality control**

We selected 96 candidate SSCX SNPs from sequences described in the study of Wiedmann et al. (2008), chosen according to their position (identified in the PAR) and to fulfill Illumina design requirements. The selected SNPs were genotyped using a Veracode Golden Gate Genotyping Assay Kit and analyzed in a Bead Xpress Reader (Illumina, Inc.). All genotypes were assigned using GENOME STUDIO software (Illumina, Inc.) and subsequently checked manually. Database pruning was conducted with PLINK (Purcell et al. 2007), excluding those SNPs with call frequency <90% and minimum allele frequency lower than 0.01 as well as those individuals with >10% missing genotypes. SNPs were annotated using Ensembl’s biomart tool in the Sscrofa10 assembly (http://www.biomart.org/).

**Statistical analysis**

With the aim of localizing the PARs and NPARs, observed (Ho) and expected (He) heterozygosities were obtained for each locus within sex using ARLEQUIN 3.5 software (Excoffier & Lischer 2010). Successive analyses were conducted for the PARs and the NPARs independently.

To test the hypothesis of genetic structuring in this sample assay, populations were grouped into seven meta-populations according to genetic, historic, and geographic relationships: Asia, Asian wild boar, local European breeds (Europe), European wild boar (which includes Tunisian wild boar), Africa, and American creole and feral pigs (Ramirez et al. 2009). Analysis of molecular variance (AMOVA) implemented in ARLEQUIN software was used to measure the degree of structuring between and within meta-populations. In males, we converted the NPAR haploid genotypes into diploid homozygous genotypes (Casto et al. 2010). We also explored alternative approaches such as reconstructing female haplotypes and diploidizing each, but the results did not change: P-values were calculated by performing 10 000 permutations.

Two classification approaches, supervised and unsupervised, were performed to characterize genetic relationships between populations. First, clustering of populations was performed with discriminant analysis of principal components (DAPC) implemented in the ADEGENET package (Jombart 2008). Second, probabilistic assignment to K groups with a Bayesian method in STRUCTURE software (Pritchard et al. 2000) was used. In this analysis, two population structures were evaluated: the first including the seven meta-populations and the second considering the 53 populations (country–breeds combination) independently. For each population structure, 10 simulations with K values ranging from 2 to 12, considering an admixture model and allele frequencies correlated, were evaluated. For each simulation, a burn-in period of 50 000 iterations was followed by 500 000 final iterations. To infer the optimal K value, STRUCTURE results were analyzed according to the delta K method described by Evanno et al. (2005).
**Results**

Quality control

A total of 56 SNPs (Table S2) passed quality control tests and were included in the analysis. The ratio of successfully genotyped SNPs (56/96 = 0.58) was much lower than that expected with this technology. The main cause of SNP discard was the high number of missing values per sample, but we also found nine non-segregating SNPs. Data are available in PLINK format upon request.

Localization of the presumable PAR in SSCX

To estimate the localization of the PAR, heterozygosity for each SNP was calculated within sex. The first 33 SNPs, located between the positions of 1.51 and 6.77 Mb, showed heterozygosities higher than zero in males (\(H_0 = 0.21\)) and comparable to those in females (\(H_0 = 0.20\)), whereas the remaining 23 SNPs, localized between 7.86 and 116.43 Mb, showed values equal to zero. In contrast, females showed \(H_0\) values higher than zero for those SNPs (\(H_0 = 0.15\)). This suggests that the PAR likely is located between 0 and 7.8 Mb in the SSCX. The PAR and NPAR were analyzed separately.

Allelic frequencies and population structure

The heterozygosities in the PAR and NPAR for each of the seven meta-populations are shown in Table 1. Overall Africa, commercial and American populations showed the highest values of \(H_0\) in the PAR and \(H_e\) in NPAR. However, these also were the most extensively sampled populations. In contrast, local breeds and wild boars from Europe showed the lowest \(H_0\) and \(H_e\). This is in agreement with previous results (e.g., Ramirez et al. 2009), which also described low variability levels in European pigs, especially in wild boars. Also, it can be noticed that the NPAR heterozygosities are lower than that of the PAR, except in European local pigs. Nevertheless, the reduction in heterozygosity is lower than expected, that is, 75%.

To gain further insight into population structuring and the effects of each potential structuring level (i.e., breed, country, and chromosome region), we performed several AMOVAs (Table 2). First, we considered all breed–country populations and second, the seven meta-populations described; finally, we analyzed each meta-population individually. Irrespective of classification, the same trends were consistently observed: the \(F_{ST}\) values were higher in the NPAR than in the PAR, and the among-population variance component also was higher in the NPAR than in the PAR—even if the within-population component dominates (Table 2). Population structure analyses suggest that 29.7 and 40.5% of the genetic variance was explained by the among-population variance component in the PAR and the NPAR, respectively, when all breed–country groups were considered (Table 2). When pigs were grouped by each of the seven meta-populations, the among-population variance component decreased, explaining only 22.7% and 25.0% of the total variance in the PAR and the NPAR, respectively. This suggests that the meta-population classification was less biologically meaningful than the breed–country arrangement.

Pairwise \(F_{ST}\) values among the seven meta-populations showed high differentiation in both the PAR and the NPAR between Asian and other populations (Table 3). Furthermore, pairwise \(F_{ST}\) values suggest that each SSCX region tells different, albeit similar, stories about the relationship between populations. In agreement with previous results (Ramirez et al. 2009), we found very low differentiation between local European breeds and wild boars; yet again, it was higher in the NPAR than in the PAR, 0.06 vs. 0.05, respectively. In contrast, differentiation between Asian domestic and wild boars was much higher, especially in the NPAR (\(F_{ST} = 0.60\)). As for derived African and American populations, our analyses indicate that, on average, the closest related breeds were the commercial breeds, followed by the European local breeds. This result agrees with a primary Iberian origin of the American populations followed by an important international breed introgression that has blurred the initial Iberian origin of American creole pigs.

<table>
<thead>
<tr>
<th>Population</th>
<th>n</th>
<th>%NA</th>
<th>(H_0) ± 0.22</th>
<th>(H_e) ± 0.19</th>
<th>(H_{E_{N P A R}}) ± 0.21</th>
<th>(H_{E_{P A R}}) ± 0.21</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asia</td>
<td>6</td>
<td>1.48</td>
<td>0.17 ± 0.22</td>
<td>0.27 ± 0.19</td>
<td>0.30 ± 0.21</td>
<td>1.11</td>
</tr>
<tr>
<td>Asia WB(^1)</td>
<td>37</td>
<td>2.99</td>
<td>0.14 ± 0.16</td>
<td>0.21 ± 0.19</td>
<td>0.15 ± 0.17</td>
<td>0.71</td>
</tr>
<tr>
<td>Europe local</td>
<td>19</td>
<td>0.75</td>
<td>0.14 ± 0.12</td>
<td>0.17 ± 0.15</td>
<td>0.16 ± 0.20</td>
<td>0.94</td>
</tr>
<tr>
<td>Europe WB</td>
<td>42</td>
<td>1.19</td>
<td>0.14 ± 0.17</td>
<td>0.16 ± 0.19</td>
<td>0.14 ± 0.20</td>
<td>0.87</td>
</tr>
<tr>
<td>Commercial</td>
<td>66</td>
<td>0.81</td>
<td>0.28 ± 0.11</td>
<td>0.36 ± 0.11</td>
<td>0.25 ± 0.20</td>
<td>0.69</td>
</tr>
<tr>
<td>Africa</td>
<td>28</td>
<td>1.53</td>
<td>0.26 ± 0.15</td>
<td>0.31 ± 0.14</td>
<td>0.25 ± 0.19</td>
<td>0.80</td>
</tr>
<tr>
<td>America</td>
<td>114</td>
<td>1.20</td>
<td>0.21 ± 0.13</td>
<td>0.25 ± 0.15</td>
<td>0.20 ± 0.20</td>
<td>0.80</td>
</tr>
<tr>
<td>Total</td>
<td>312</td>
<td>1.33</td>
<td>0.21 ± 0.10</td>
<td>0.32 ± 0.13</td>
<td>0.25 ± 0.18</td>
<td>0.78</td>
</tr>
</tbody>
</table>

\(^1\)WB, wild boar.
Probabilistic assignation to genetic groups

Differentiation of Asian populations from other groups was confirmed with DAPC in both SSCX regions (Fig. 1). Principal components retained explained 91% of the genetic variance in both the PAR and NPAR. This analysis (Fig. 1) corroborates the similarity in allelic frequencies between European local pigs and wild boars and also among the non-Asian populations. Asian local populations showed a cline-like relationship between Asian wild boars and commercial populations in the PAR (Fig. 1a). However, the discriminant analysis was more difficult to interpret regarding the NPAR (Fig. 1b). In this case, DAPC showed a higher similarity of European and commercial pigs with Asian wild boars than with Asian domestic pigs. We hypothesize that demographic effects in Asian domestication together with ascertainment bias effect and limited sampling could explain these results.

We also performed an unsupervised Bayesian probabilistic assignation of individuals to genetic groups, as implemented in structure software. The most likely number of clusters was \( K = 2 \) when pigs were grouped by meta-population. In agreement with numerous previous results, Asian populations clustered in a separate group from the remaining populations (Fig. 2). Moreover, European domestic and wild boar pigs clustered in a second genetic group with a more than 97% membership proportion. In addition, commercial, African and American populations showed different degrees of admixture of European and Asian genetic components, the latter genetic component being in lower proportion.

To assess structuring in more detail, we also ran structure with pigs grouped by breed and country. Here, \( K = 3 \) was the most supported value. Figure 3 shows the average cluster membership of pigs by breed and by chromosome.

### Table 2 AMOVA results for the different population structures in each SSCX region.

<table>
<thead>
<tr>
<th>Group</th>
<th>Populations</th>
<th>Pseudoautosomal region</th>
<th>Non-pseudoautosomal region</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Within populations</td>
<td>Among populations</td>
</tr>
<tr>
<td>Breed-country</td>
<td>53</td>
<td>3.26</td>
<td>1.37</td>
</tr>
<tr>
<td>Meta-populations</td>
<td>7</td>
<td>3.89</td>
<td>1.14</td>
</tr>
<tr>
<td>Asia WB</td>
<td>5</td>
<td>2.31</td>
<td>0.54</td>
</tr>
<tr>
<td>Europe</td>
<td>5</td>
<td>2.38</td>
<td>0.24</td>
</tr>
<tr>
<td>Europe WB</td>
<td>8</td>
<td>2.33</td>
<td>0.42</td>
</tr>
<tr>
<td>Commercial</td>
<td>16</td>
<td>4.83</td>
<td>0.76</td>
</tr>
<tr>
<td>Africa</td>
<td>5</td>
<td>4.54</td>
<td>0.31</td>
</tr>
<tr>
<td>America</td>
<td>13</td>
<td>3.01</td>
<td>0.60</td>
</tr>
</tbody>
</table>

\(^1\)Includes the seven meta-populations Asia, Asia wild boar, Europe, Europe wild boar, Commercial, Africa, and Americas.

\(^2\)Asia was excluded because all individuals were considered one population.

\(^3\)WB, wild boar.

\(^4\)No significant statistical differences were found. The rest of the \( F_{ST} \) values were highly significant.
region. Overall, Asian and European wild boars and Chinese pigs were rather homogeneous, especially for the PAR. Interestingly, Japanese wild boars from Ryukyu and the main islands were slightly different in the NPAR, in parallel with results from mtDNA (Wu et al. 2007) which suggest different founder origins. The native Korean pigs showed differences in allelic frequencies to commercial breed introgression signals. In fact, the origin of native Korean pigs is mixed, with accredited introgression of commercial pigs within the original Korean germplasm. Interestingly, the composition of the PAR and NPAR was quite distinct, with a larger proportion of European clusters in the NPAR, again in agreement with a primary European origin via males.

European populations were made up of two genetic clusters in different proportions depending on the SSCX region. In the PAR, one cluster represented 80.7% of the

![Figure 2](image1.png)

**Figure 2** Bayesian probabilistic assignation of individuals considering meta-population structure in the pseudoautosomal regions (a) and the non-pseudoautosomal region (b). Breeds included in each group are described in Table S1.

![Figure 3](image2.png)

**Figure 3** Average Bayesian probabilistic cluster assignation by breed and country in the pseudoautosomal regions (a) and the non-pseudoautosomal region (b). Breed and country codes as in Table S1.
genetic component in wild boars; this value decreased to 69.3% in the NPAR, influenced by high differences in allelic frequencies of Italian and Armenian individuals. Mangalitsa pigs showed allelic frequencies similar to that of Iberian pigs in the PAR, whereas in the NPAR, they were similar to Armenian pigs. As expected, commercial breeds were mosaics of European and Asian-like clusters in different degrees. However, there were differences between the PAR and NPAR, maybe as a result of differences in the sex ratio of the original breed genetic contribution to each population. For instance, in Duroc and in Pietrain, the Asian-like component decreased in the NPAR. Furthermore, clustering was very different between the PAR and NPAR for breeds like Tamworth, Large Black, and Chester White. As in commercial breeds, three clusters with different proportions for each SSCX region were detected in America populations. Yucatan, Feral Argentinean, and Peruvian pigs were primarily European in origin, whereas an Asian-like component appeared in the NPAR for Brazilian Monteiro and Cuban pigs. In a previous work on the Y chromosome in African populations, we observed a larger Asian component in Eastern than in Western countries; however, this trend is not apparent from our results for SSCX.

**Discussion**

This study is a first approach to evaluate the usefulness of SNP polymorphisms in the pig X chromosome to detect population structure in a worldwide pig sample. Of 96 candidate SNPs, we retained only 56 polymorphic SNPs. These SNPs were initially obtained from pools of seven pig breeds, namely Duroc, Landrace, Yorkshire, Large White, Hampshire, Berkshire, and Pietrain, so they represent a limited variability in the whole species (Wiedmann et al. 2008).

Using indirect evidence of male heterozygosity, we determined that the PAR spans the first 7.8 Mb of the SSCX short arm. Quilter et al. (2002) mapped the STS, KAL1, and PRKX genes to the PAR using fluorescent in situ hybridization (FISH); in the Sscrofa9 assembly, the first two genes are located at 2.89 and 4.00 Mb, which does suggest a bigger PAR length in pig than in humans although smaller than in cattle (Ross et al. 2005; Das et al. 2009). Intriguingly, gene PRKX (ENSSSCG00000012833) is located on the Xq tail at 125.63–125.80 Mb. Further, Rohrer et al. (1994) reported three markers presumably located in the PAR; one of the markers, namely SW949, was aligned against Sscrofa9 assembly and matched two positions: 125.67 (P-value 1.8e-146) and 125.77 Mb (P-value 6.3e-144), again on the Xq tail. These data point to possible errors in the X chromosome Sscrofa9 assembly, given that a single PAR is suggested by FISH data.

The SHROOM2 gene has been associated with PABs in pigs (Raussepp et al. 2012). In the Sscrofa9 assembly, SHROOM2 (ENSSSCG00000012104) is located at 5.26 Mb and within the bounds of the PAR suggested in this study. The amelogenin (AMEL) gene has been reported as the ancient Pseudoautosomal Boundary (PAB) in mammals (Iwase et al. 2003). This gene is located between 6.89 and 6.90 Mb in the pig genome (ENSSSCG00000012113), and male heterozygosity was zero in this region. Therefore, a comparative map of PAR in humans, cattle, goat, sheep, horse, and dog (Das et al. 2009), and the results reported here suggest that the PAR size in pigs is ~7 Mb.

The main feature of the X chromosome in population genetics studies is its low effective size, which theoretically reduces the genetic variability to three-quarters that of autosomes. Our data do show a reduction in the variability in the NPAR vs. PAR (Table 1), but this was much smaller than expected, except in commercial breeds—where the SNPs were ascertained. Although deviations from theoretical variability ratio have been reported (e.g., Gottipati et al. 2011), the most likely explanation for our results is SNP ascertainment bias. The analyses from the few published sequence data suggest that, in pigs, the ratio of X/A variability is actually much lower than 0.75 (Amaral et al. 2011; Esteve-Codina et al. 2011). Moreover, Ma et al. (2010) found a large (~31 Mb) recombination cold spot adjacent to the centromere of the pig X chromosome. Here, we found a strong reduction in Ho values in SNPs located between 61.52 and 92.32 Mb, in a position similar to that

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**Table 3** Pairwise $F_{ST}$ values in pseudoautosomal regions (below diagonal) and non-pseudoautosomal region (above diagonal).

<table>
<thead>
<tr>
<th></th>
<th>Africa</th>
<th>America</th>
<th>Asia</th>
<th>Asia WB</th>
<th>Europe</th>
<th>Europe WB</th>
<th>Commercial</th>
</tr>
</thead>
<tbody>
<tr>
<td>Africa</td>
<td>0.10</td>
<td>0.52</td>
<td>0.27</td>
<td>0.16</td>
<td>0.21</td>
<td>0.02</td>
<td></td>
</tr>
<tr>
<td>America</td>
<td>0.03</td>
<td>0.64</td>
<td>0.45</td>
<td>0.05</td>
<td>0.15</td>
<td>0.04</td>
<td></td>
</tr>
<tr>
<td>Asia</td>
<td>0.18</td>
<td>0.28</td>
<td>0.60</td>
<td>0.64</td>
<td>0.69</td>
<td>0.57</td>
<td></td>
</tr>
<tr>
<td>Asia WB</td>
<td>0.44</td>
<td>0.50</td>
<td>0.22</td>
<td>0.51</td>
<td>0.53</td>
<td>0.34</td>
<td></td>
</tr>
<tr>
<td>Europe</td>
<td>0.05</td>
<td>0.03</td>
<td>0.36</td>
<td>0.58</td>
<td>0.06</td>
<td>0.07</td>
<td></td>
</tr>
<tr>
<td>Europe WB</td>
<td>0.09</td>
<td>0.08</td>
<td>0.41</td>
<td>0.61</td>
<td>0.05</td>
<td>0.13</td>
<td></td>
</tr>
<tr>
<td>Commercial</td>
<td>0.02</td>
<td>0.06</td>
<td>0.11</td>
<td>0.32</td>
<td>0.10</td>
<td>0.15</td>
<td></td>
</tr>
</tbody>
</table>

$^1$WB, wild boar.

All $F_{ST}$ values were significant ($P < 0.05$).
in the study of Ma et al. Detailed analyses for this region showed a single haplotype in all samples except for two native Korean sows.

Overall, commercial populations showed the highest Ho and He values, followed by African and American populations. Given that the populations from these two continents are derived, a lower variability should be expected; these data then would suggest an important introgression of commercial breeds into the ‘creole’ populations. In general, Ho and He values were higher in Asian than in European populations, as reported in other studies using autosomal microsatellite markers (Fan et al. 2002; Luetkemeier et al. 2010). Moreover, as also reported by Ramirez et al. (2009), both domestic and European wild boar populations showed similar heterozygosity values for both SSCX regions and low differentiation, suggesting a possible gene flow between populations. In Asia, Ho and He values were higher in domestic populations than in wild boar, in contrast to results from Luetkemeier et al. (2010). However, these Ho and He values should be interpreted with caution, given the small sample size of some populations and differences in allele frequencies with commercial populations (ascertainment bias effect).

The X chromosome patterns of variability are strongly influenced by the sex ratio: genetic drift is very sensitive to the number of males, whereas the recombination rate depends on that of females. As revealed by STRUCTURE, a variety of different patterns between the PAR and NPAR, especially in commercial breeds, can be seen (Fig. 3). In some breeds, such as Berkshire or British Lop and most wild boars, the PAR and NPAR exhibited similar cluster compositions, but in most breeds, there were large differences between the PAR and NPAR, showing that their genetic trajectories are distinct.

Discriminant analysis of principal components and FST results showed a low differentiation between Africa, America, and commercial populations. Historical records document a strong international breed introgression in Africa, especially from the maternal side, from Asia and British breeds during the 18th and 19th centuries. Ramirez et al. (2009) found a primary European influence in western Africa, whereas eastern Africa exhibited an Asian genetic component. Our results cannot confirm these results; Eastern African populations (Kenya, Tanzania, and Uganda) share a large proportion of a European-like genetic group in both SSCX regions (Fig. 3). American population origins are European, mainly Iberian, with recent commercial breed introgression (Ramirez et al. 2009; Souza et al. 2009). Our results are consistent with this, showing low differentiation with the commercial population; in addition, all breeds showed a proportion of an Asian-like cluster (Fig. 3). Some populations (Yucatan, Argentinean feral pigs, and Guatemalan pigs) still preserve a high proportion of a European-like genetic cluster proportion in both regions, whereas others (Costa Rican or Argentinean creoles) show strong influences of a commercial-like genetic component. A particularly interesting case was found in the Brazilian Moura and Monteiro breeds, where the Asian-like component appears in both populations but in different SSCX regions. The origin of the Monteiro breed is possibly European but has been crossbred with Asian breeds to improve reproductive parameters (Grossi et al. 2006). Overall, it is difficult to quantify the introgression impact of commercial breeds in American populations.

Conclusions

Thus far, the X chromosome has been poorly studied in the pig species. In this genotyping study, our findings are in agreement with a reduction in the variability in the NPAR, although attenuated because of SNP ascertainment bias. We also find indirect evidence that the PAR comprises approximately ~7 Mb of SSCX. Two main population clusters were detected, corresponding to Asian and European origins. Nevertheless, our results show that genetic relationships between populations can vary greatly between the NPAR and the PAR, underscoring the fact that their genetic trajectories can be quite different. The NPAR showed an increased commercial-like genetic component relative to the PAR, probably because of the fact that human selection processes to obtain individuals with high productive parameters were mediated by introgressing boars rather than sows. Further studies with a much denser SNP panel should allow the detection of selective sweeps in this important chromosome.

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References

tion revealed through massive parallel sequencing of pooled DNA. PLoS ONE 6, e14782.


Supporting information

Additional supporting information may be found in the online version of this article.

Table S1 Breed codes and ISO country codes employed and breed x country sample size.

Table S2 Submission name, dbSNP accession number and position on X chromosome for each SNP.

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